

Research

Open Access

Effects of *Fructus Piperis Longi* extract on fibrotic liver of gamma-irradiated rats

Somaya Zakaria Mansour*¹ and Hanan El-Kabany²

Address: ¹Radiation Biology Department, National Centre for Radiation Research and Technology, Atomic Authority, Cairo, Egypt and ²Health Radiation Research Department, National Centre for Radiation Research and Technology, Atomic Authority, Cairo, Egypt

Email: Somaya Zakaria Mansour* - szmansour@yahoo.com; Hanan El-Kabany - moonyhabiby@hotmail.com

* Corresponding author

Published: 30 January 2009

Received: 7 April 2008

Chinese Medicine 2009, 4:2 doi:10.1186/1749-8546-4-2

Accepted: 30 January 2009

This article is available from: <http://www.cmjournal.org/content/4/1/2>

© 2009 Mansour and El-Kabany; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: A major biomarker for liver fibrosis is transglutaminase which catalyzes cross-linking of epsilon-amines and alpha-glutamyl residues among amino acids leading to fibrosis. *Fructus Piperis Longi* is a common herb used in Chinese medicine. The present study evaluates the role of the ethanol extract of *Fructus Piperis Longi* in the modulation of liver function in liver fibrosis.

Methods: Plf extract (50 mg/kg) was force-fed to rats every other day 7 days before administration of thioacetamide and/or gamma irradiation. Thioacetamid 200 mg/kg was intraperitoneally administered to rats twice per week for four weeks. Rats were gamma irradiated (2 Gy/week up to a total dose of 8 Gy). Administration of Plf ext was extended during thioacetamid and/or irradiation treatment. Animals were sacrificed. Biochemical parameters in homogenised liver were tested.

Results: A significant increase in transglutaminase activity and collagen content was recorded in the liver of thioacetamid-treated and/or irradiated rats. Significant increases in lipid peroxides, lipid hydroperoxides and conjugated dienes associated to significant decreases of reduced glutathione content, superoxide dismutase and catalase activities were also recorded. Administration of Plf ext treatment reduced the severity of liver fibrosis and oxidative damage which was substantiated by amelioration of liver function detected by a decrease in serum aspartate aminotransaminase, alanine aminotransferase, alkaline phosphatase, gamma glutamyltransferase activities and bilirubin (total, direct and indirect) content.

Conclusion: Treatment of the ethanolic extract of *Fructus Piperis Longi* ameliorated the increase of the activity of tTG enzyme and enhanced antioxidant activities in fibrotic liver.

Background

Fibrosis of the liver is a state of complicated end stage alteration of structure and function due to different aetiologies. Fibrosis is a consequence of different prevalent mechanisms according to the diverse causes of parenchymal damage. Fibrosis caused by chronic viral infection is

initially concentrated within and around the portal tract, while fibrosis secondary to toxic/metabolic damage is located mainly in the centrolobular areas [1].

Oxidative stress, characterized by the overproduction of reactive oxygen species (ROS), which overwhelm the lev-

els of antioxidants, has been suggested as the pathogenic factor of a number of human diseases and was reported to cause tissue damage [2]. ROS can react with cellular macromolecules such as nucleic acids, polyunsaturated fatty acids in cellular membranes and sulfhydryl bonds in proteins to cause mutagenesis, carcinogenesis and cell death.

Thioacetamide (TAA, $\text{CH}_3\text{-C[S] NH}_2$), a known fungicide used to control the decay of fruits [3] was shown to be S-oxidized at the thioamide group to TAA sulfoxide ($\text{CH}_3\text{-C[SO] NH}_2$) and subsequently di-Sulfoxide ($\text{CH}_3\text{-C[SO}_2\text{] NH}_2$) in the liver. The reactive intermediates in this pathway covalently bind to hepatic macromolecules and eventually cause liver injury [4,5], whereby free radical-mediated lipid peroxidation contributes to the development of TAA induced liver fibrosis [6,7]. Prolonged administration of TAA causes hyperplastic liver nodules, liver cell adenomas and hepatocarcinomas. The free radicals produced during TAA metabolism interfere with ribosomal activity, thereby hindering protein synthesis [8]. The biochemical and morphological changes observed in TAA-induced rat liver injury resemble to a large extent human liver disease and could serve as a suitable model for studying the causes of human liver fibrosis and cirrhosis [9].

Tissue fibrosis is associated with the increase of the tTG activity and accumulation of ECM [10]. In liver fibrosis induced in rats by carbon tetrachloride (CCl_4) and in human patients with an acute liver disease, Mirza *et al.* [11] found a dramatic rise in tissue transglutaminases (tTG) activity. The enzyme catalyzes the specific cross-linking of ϵ -amines and α -glutamyl residues among amino acids [12]. This activity leads to the cross-linking of extracellular matrix (ECM) proteins thereby increasing the deposition [13] of such proteins and their resistance to proteolytic enzymes, which leads to tissue fibrosis [14,15]. Several studies specifically described the role of tTG in cross-linking of fibronectin, osteonectin, osteopontin, laminin and other extracellular matrix components [12].

The pathogenesis of liver fibrosis is not clear; however, it was suggested that an increase of ROS coupled with a decrease in body antioxidant system activity play an important role in the pathological changes, particularly in the cases of radiation exposure and liver toxicity [16]. Radiation exposure may cause disruption of normal cell membranes as a result of direct interaction of radiation with cellular membranes or through the action of free radicals produced by radiation [17].

Several endogenous protective mechanisms may limit ROS and the damage caused by them [18]. However, this protection may be insufficient. When the formation of ROS is excessive, additional protective mechanisms of die-

tary, antioxidants may help maintain liver functions. Several natural antioxidants were proposed to prevent and treat hepatopathies induced by oxidative stress [19]. Rich in flavonoids, *Fructus Piperis Longi* (Bibo, long pepper) is used in Chinese medicine to treat various conditions such as jaundice and allergy [20]. It has been demonstrated to be anti-tussive, anti-asthmatic, anti-allergic, anti-tubercular, antipyretic, hypotensive, hypoglycemic, antihelminthic and coronary vasodilatory [21]. The aim of the present study is to investigate the hepatoprotective activity of *Fructus Piperis Longi* against liver fibrosis.

Methods

Materials and instrument

Fructus Piperis Longi was obtained from the local market. All chemicals and reagents used in the experiment were of analytical grade and purchased from either Merck (Germany) or Sigma Aldrich Chemie (Germany). Assay kits for testing alkaline phosphatase (ALP), alanine aminotransferase (ALT), aspartate aminotransaminase (AST), gamma glutamyltransferase (GGT), bilirubin and total protein were supplied by Diamond Diagnostics (Egypt).

Instrument included electric digital balance (Shimadzu, Type Ay 220, Japan), pH meter (Jenway, UK), homogenizer (Glas-col, TERRE HAUT, USA), cooling centrifuge (Mettler, Model K23, Germany), centrifuge (Janetzi, Model T30, Germany), shaker incubator (Lab-Line Instruments, USA) and spectrophotometer (Helios, UV/Visible, UK).

Experimental animals

All animal treatment procedures conformed to the National Institutes of Health (NIH) guidelines [22]. Sprague Dawley male albino rats (170–220 g) were used in this study. Animals were obtained from the National Centre for Radiation Research and Technology (NCCRT), Cairo, Egypt. The animals were housed in cages and maintained under standard conditions of ventilation, temperature and humidity. Animals received standard food pellets and water *ad libitum*.

Gamma irradiation procedure

Irradiation of animals was carried out at the National Centre for Radiation Research and Technology (NCRRT) in Cairo, Egypt, with a Gamma cell-40 (Cesium-137 irradiation units, Canada). The irradiation dose rate was 0.61 Gy/min. Animals (whole body) were exposed to 2 Gy per week at a total dose of 8 Gy, one day after TAA administration.

Induction of liver fibrosis

Liver fibrosis was induced by intraperitoneal administration of 200 mg/kg TAA twice per week for four weeks according to El Borai *et al.* [23].

Preparation of *Fructus Piperis Longi* extract

The ethanolic extract of *Fructus Piperis Longi* was prepared according to Christina *et al.* [21]. *Fructus Piperis Longi* was obtained from the local market and was dried and powdered. About 500 g of dry powder was extracted with 5L of ethanol at 60–70 °C for 72 hours by continuous hot percolation with a Soxhlet apparatus. The ethanolic extract was then filtered and concentrated by vacuum distillation to dry. The yield for 500 g was 37 g. This dried extract was then stored at 4 °C until use. Rats were force-fed 50 mg/kg of distilled water per day for five weeks starting from seven days before TAA administration.

Animal groups

The experimental animals were divided into eight groups ($n = 6$), namely (1) Control: healthy animals received distilled water; (2) Plf ext: animals received *Fructus Piperis Longi* extract; (3) TAA: animals were injected with TAA; (4) Plf ext + TAA: animals received *Fructus Piperis Longi* extract and were injected with TAA; (5) γ irradiation: animals were exposed to γ irradiation; (6) Plf ext + γ irradiation: animals received *Fructus Piperis Longi* extract and were exposed to γ irradiation; (7) TAA + γ irradiation: animals were injected with TAA and exposed to γ irradiation and (8) Plf ext + TAA + γ irradiation: animals received *Fructus Piperis Longi* extract and were injected with TAA and exposed to γ irradiation.

Rats of all groups received the last irradiation exposure on the day before overnight fasting and sacrifice. Blood samples were collected by heart puncture. Plasma of each blood sample was separated and kept frozen for biochemical assays. Liver samples were kept at -80 °C until biochemical assays. Liver tissue homogenate (10% w/v) in phosphate-buffered-saline (0.02 M sodium phosphate buffer with 0.15 M sodium chloride, pH7.4) was prepared with a portion of liver homogenized in a glass tissue homogenizer with a Teflon pestle.

Biochemical assays

Tissue transglutaminase (tTG) activity in 100 μ l liver homogenate was determined according to the direct spectrophotometric method by De Macedo *et al.* [24]. Total protein content in liver tissue was determined according to the method by Henry [25] to calculate the specific enzyme activity of tTG in the liver. Liver collagen content was determined according to Woessner [26].

Reduced glutathione concentration (GSH) in liver was determined according to Beutler *et al.* [27]. Superoxide dismutase (SOD) activity in liver was measured according to Minami and Yoshikawa [28]. The colorimetric assay for liver catalase activity (Cat) was carried out according to Sinha [29]. Lipid peroxides (LP) indicated by the formation of malondialdehyde (MDA) was assessed in liver

homogenates according to Yoshioka *et al.* [30]. Lipid hydroperoxides (LHP) in liver was determined according to the Fox method described by Jiang *et al.* [31]. The levels of conjugated dienes (CD) in liver were measured according to Rechnagel and Gglende and Nowak *et al.* [32,33].

Plasma ALP activity was determined according to Teitz [34]. Activities of ALT and AST were determined colorimetrically according to Reitman and Frankel [35]. Plasma gamma GGT activity was measured kinetically according to Szasz [36]. Plasma bilirubin (total, direct and indirect) contents were determined according to Perry *et al.* [37].

Statistical analysis

The SPSS (version 10) was used in data analysis. Data were analyzed with one-way analysis of variance (ANOVA) followed by a *post hoc* test (LSD alpha) for multiple comparisons. The data were expressed as mean \pm standard deviation (SD). P values < 0.05 were considered to be statistically significant.

Results

Administration of *Fructus Piperis Longi* ethanol extract (Plf ext) to rats, by force-feeding, for a period of five weeks, did not show significant changes in all the studied parameters, indicating that the extract did not affect the liver functions (Tables 1, 2, 3, 4, 5).

As shown in Table 1, TAA significantly increased ($P = 0.0001$) liver collagen content and tTG activity. Irradiated rats showed significantly increased liver collagen content ($P = 0.019$) and tTG activity ($P = 0.0001$). In the TAA + irradiation group, significant increases ($P = 0.0001$) in liver collagen content and tTG activity were observed. Treatment of Plf ext significantly ameliorated ($P = 0.0001$, no significance, 0.0001, 0.0001, 0.0001 and 0.0001 respectively) the increase of collagen content and tTG activities in the rats that received TAA or γ -irradiation or both (Table 1).

TAA induced significant decreases ($P = 0.0001$) in liver GSH content, SOD and Cat activities (Table 2), which were parallel to significant increases in LP ($P = 0.0001$), LPH ($P = 0.003$) and CD ($P = 0.0001$) content (Table 3). Irradiated rats showed significant decreases ($P = 0.0001$) in liver GSH content and Cat activity in association with significant increases in LP ($P = 0.0001$), LPH ($P = 0.008$) and CD ($P = 0.002$) content (Tables 2 and 3). In the TAA + irradiation group, significant decreases ($P = 0.0001$) in liver GSH content, SOD and Cat activities in association with significant increases in LP ($P = 0.0001$), LPH ($P = 0.001$) and CD ($P = 0.0001$) content were observed. Treatment of Plf ext significantly reduced ($P = 0.0001$) oxidative stress in the rats that received TAA or γ -irradiation or both.

Table 1: Transglutaminase (tTG) activity and collagen content in liver tissue homogenates of rats under different treatment conditions

Groups	tTG (anilide/umol/mg protein/min)	Collagen (mg/g wet tissue)
Control	1.26 ± 0.182	3.96 ± 0.346
Pif ext	1.32 ± 0.111	4.03 ± 0.221
TAA	2.87 ± 0.166	6.14 ± 0.318
a	P = 0.0001	P = 0.0001
Pif ext+TAA	1.58 ± 0.085	4.60 ± 0.163
a	P = 0.007	P = 0.013
b	P = 0.0001	P = 0.0001
γ radiation	2.26 ± 0.070	4.56 ± 0.253
a	P = 0.0001	P = 0.019
Pif ext+γ radiation	1.37 ± 0.082	4.20 ± 0.499
a	NS	NS
c	P = 0.0001	NS
TAA+γ radiation	3.49 ± 0.185	6.77 ± 0.075
a	P = 0.0001	P = 0.0001
b	P = 0.0001	P = 0.015
c	P = 0.0001	P = 0.0001
Pif ext+TAA+γ radiation	1.76 ± 0.065	5.13 ± 0.125
a	P = 0.0001	P = 0.0001
b	P = 0.0001	P = 0.0001
c	P = 0.0001	P = 0.023
d	P = 0.0001	P = 0.0001

Each value represents mean ± SD of 6 determinations.

a: significance over control group

b: significance over TAA group

c: significance over γ radiation group

d: significance over TAA+γ radiation group

NS: no significance

Table 2: SOD and Cat activities and GSH content in liver tissue homogenates of rats under different treatment conditions

Groups	SOD (μg/g wet tissue)	Cat (μmol/g wet tissue)	GSH (mg/g wet tissue)
Control	12.06 ± 0.701	119.36 ± 9.799	24.75 ± 1.035
Pif ext	12.38 ± 0.499	119.37 ± 5.334	25.06 ± 0.752
TAA	10.79 ± 0.054	101.40 ± 3.561	21.02 ± 0.699
a	P = 0.0001	P = 0.0001	P = 0.0001
Pif ext+TAA	11.84 ± 0.398	118.89 ± 4.522	23.26 ± 0.586
a	NS	NS	P = 0.0001
b	NS	P = 0.001	P = 0.0001
γ radiation	11.52 ± 0.151	99.63 ± 9.279	20.60 ± 0.793
a	NS	P = 0.0001	P = 0.0001
Pif ext+γ radiation	11.88 ± 0.031	117.32 ± 9.122	23.11 ± 0.867
a	NS	NS	P = 0.0001
c	NS	P = 0.001	P = 0.0001
TAA+γ radiation	10.64 ± 0.316	91.01 ± 6.793	19.86 ± 0.263
a	P = 0.0001	P = 0.0001	P = 0.0001
b	NS	P = 0.041	P = 0.009
c	P = 0.002	NS	NS
Pif ext+TAA+γ radiation	11.82 ± 0.880	111.82 ± 14.453	22.80 ± 0.594
a	P = 0.001	NS	P = 0.0001
b	P = 0.001	P = 0.041	P = 0.0001
c	NS	P = 0.018	P = 0.0001
d	P = 0.0001	P = 0.0001	P = 0.0001

Each value represents mean ± SD of 6 determinations.

a: significance over control group

b: significance over TAA group

c: significance over γ radiation group

d: significance over TAA+γ radiation group

NS: no significance

Table 3: Lipid peroxides (LP), lipid hydroperoxide (LHP) and conjugated diene (CD) in liver tissue homogenates of rats under different treatment conditions

Groups	LP (µg/g tissue)	LHP (µM)	CD (Abs ₂₃₄ /g tissue)
Control	108.27 ± 7.246	17.81 ± 0.841	2.20 ± 0.091
Plf ext	114.72 ± 5.170	17.90 ± 0.759	2.20 ± 0.068
TAA	169.04 ± 16.914	21.64 ± 1.995	3.14 ± 0.042
a	P = 0.0001	P = 0.003	P = 0.0001
Plf ext+TAA	127.47 ± 2.480	17.79 ± 2.949	2.30 ± 0.200
a	P = 0.003	NS	NS
b	P = 0.0001	P = 0.003	P = 0.0001
γ radiation	135.54 ± 13.191	20.91 ± 0.833	2.54 ± 0.191
a	P = 0.0001	P = 0.008	P = 0.002
Plf ext+γ radiation	123.98 ± 4.606	17.52 ± 2.553	2.24 ± 0.043
a	P = 0.012	NS	NS
c	NS	P = 0.008	P = 0.006
TAA+γ radiation	182.93 ± 9.560	21.92 ± 1.958	3.20 ± 0.015
a	P = 0.0001	P = 0.001	P = 0.0001
b	P = 0.025	NS	NS
c	P = 0.0001	NS	P = 0.0001
Plf ext+TAA+γ radiation	142.04 ± 14.092	17.88 ± 2.247	2.47 ± 0.116
a	P = 0.0001	NS	P = 0.011
b	P = 0.0001	P = 0.003	P = 0.0001
c	NS	P = 0.01	NS
d	P = 0.0001	P = 0.001	P = 0.0001

Each value represents mean ± SD of 6 determinations.

a: significance over control group

b: significance over TAA group

c: significance over γ radiation group

d: significance over TAA+γ radiation group

NS: no significance

Significant increases (P = 0.0001) of plasma ALT, AST, ALP and GGT activities were observed in rats that received TAA or γ-irradiation or both. Treatment of Plf ext ameliorated (P = 0.0001) these increases (Table 4).

Significant increases (P = 0.0001) in the content of total, direct and indirect bilirubin were observed in the rats that received TAA or γ-irradiation or both. Treatment of Plf ext ameliorated (P = 0.0001) these increases (Table 5).

Figure 1 shows a significant increase in the liver weight of the rats that received TAA or γ-irradiation or both. Administration of Plf ext did not significantly ameliorate liver weight.

Discussion

In the present study, parameters of liver fibrosis induced by TAA with or without radiation exposure were shown as an increase of liver weight, significant increases in liver tTG activity and collagen content associated with significant decreases in GSH content, SOD and Cat activities and increases in LP, LHP and CD content. Hepatic damage was indicated by significant increases of serum AST, ALT, ALP activities and bilirubin content.

The increase in tTG activity may be attributed to the increased binding of the nuclear factor-kappaB (NF-κB) to

the NF-κB motif of the tTG promoter, where tTG gene expression increases during hepatic injury and fibrosis [38]. The concomitant increase of both hepatic collagen and tTG activity may be explained by the dual effect exerted by the NF-κB, which is induced by oxidative stress

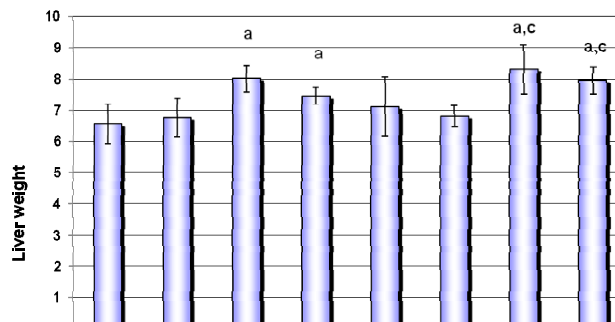


Figure 1
Rats' liver weight under various treatment conditions. Each value represents the mean ± SD of 6 determinations. a: significance of treatment over control P = 0.0001, 0.021, 0.0001 and 0.001 respectively. c: significance of treatment over γ irradiation group P = 0.003 and 0.030 respectively.

Table 4: ALT, AST, ALP and GGT activities in plasma of rats under different treatment conditions

Groups	ALT (U/l)	AST (U/l)	ALP (U/l)	GGT (U/l)
Control	15.54 ± 1.987	8.00 ± 0.503	50.11 ± 1.241	1.00 ± 0.00
Plf ext	15.44 ± 2.001	8.18 ± 0.804	49.36 ± 1.017	1.08 ± 0.144
TAA	23.58 ± 1.282	16.04 ± 3.380	94.35 ± 2.431	4.66 ± 0.212
a	P = 0.0001	P = 0.0001	P = 0.0001	P = 0.0001
Plf ext+TAA	17.03 ± 1.92	9.29 ± 0.419	54.24 ± 1.417	2.30 ± 0.282
a	NS	NS	P = 0.0001	P = 0.0001
b	P = 0.0001	P = 0.0001	P = 0.0001	P = 0.0001
γ radiation	19.91 ± 1.371	14.37 ± 0.567	88.00 ± 1.763	2.62 ± 0.247
a	P = 0.0001	P = 0.0001	P = 0.0001	P = 0.0001
Plf ext+γ radiation	17.17 ± 1.899	9.26 ± 0.585	51.98 ± 1.592	1.43 ± 0.087
a	NS	NS	NS	P = 0.036
c	P = 0.006	P = 0.0001	P = 0.0001	P = 0.0001
TAA+γ radiation	24.60 ± 1.268	21.20 ± 0.560	104.40 ± 3.241	5.78 ± 0.358
a	P = 0.0001	P = 0.0001	P = 0.0001	P = 0.0001
b	NS	P = 0.0001	P = 0.0001	P = 0.0001
c	P = 0.0001	P = 0.0001	P = 0.0001	P = 0.0001
Plf ext+TAA+γ radiation	19.85 ± 0.813	12.20 ± 1.344	55.99 ± 2.794	2.71 ± 0.276
a	P = 0.0001	P = 0.0001	P = 0.0001	P = 0.0001
b	P = 0.0001	P = 0.0001	P = 0.0001	P = 0.0001
c	NS	P = 0.011	P = 0.0001	NS
d	P = 0.0001	P = 0.0001	P = 0.0001	P = 0.0001

Each value represents mean ± SD of 6 determinations.

a: significance over control group

b: significance over TAA group

c: significance over γ radiation group

d: significance over TAA+γ radiation group

NS: no significance

[39]. Nevertheless, the association between tTG activity and fibrosis may involve other factors such as the factor-beta (TGF-β), major fibrogenic growth factors, where tTG activates the latent TGF-μ1, which in turn leads to *de novo* synthesis of tTG [40]. The increase of tTG activity may also be a consequence of GSH depletion and mitochondrial dysfunction [41].

The depletion in GSH content may be due to the oxidation of sulfhydryl group and diminished activity of glutathione reductase [42]. The significant decrease in the activity of antioxidant enzymes may be caused by cell membrane damage and alterations in dynamic permeability of membranes due to peroxidation, followed by the release of intracellular enzymes to the blood stream [43]. In addition, an excess of •OH causes oxidative damage to enzymes, resulting in the modification of their activities [43,44]. The marked increase in MDA levels is likely to be a result of the inactivation of scavenger enzymes induced by ROS [44-46]. According to Ueda *et al.* [47], the generation of lipid peroxide and its appearance in the animal's liver may be a result of a chain of reactions or may be initiated by an indirect mechanism that enables the escape from anti-oxidation.

Fructus Piperis Longi has recently been proposed as a chemopreventive agent for its antioxidant activities [48,49]. The present study showed that treatment of Plf ext significantly

reduced liver fibrosis as evidenced by significant decreases of tTG activity and collagen content, with concomitant enhancement of the antioxidant status and improvement of liver functions. The results are consistent with other reports on the role of polyphenols against oxidative stress [39] because Plf ext is rich in polyphenols [50] which may up-regulate the antioxidant [51-53], thereby decreasing the free radical-induced lipid peroxidation [51].

The increased resistance of liver tissue against liver fibrosis and oxidative stress after treatment was shown by the significant decreases in serum ALT, ALP and AST activities and bilirubin content. The results are consistent with a previous study which demonstrated that the ethanolic extract of *Fructus Piperis Longi* possessed hepatoprotective activity lowering serum enzymes ALT and AST [54]. Further studies NF-κB, tTG gene expression and TGF-B would help elucidate the mechanism of action

Conclusion

Treatment of the ethanolic extract of *Fructus Piperis Longi* ameliorated the increase of the activity of tTG enzyme and enhanced the antioxidant activities in fibrotic liver.

Abbreviations

tTG: transglutaminase; Plf ext: *Fructus Piperis Longi* ethanol extract; TAA: thioacetamide; LP: lipid peroxides; LHP:

Table 5: Bilirubin total, direct and indirect concentration in plasma of rats under different treatment conditions

Groups	Bilirubin total (mg/ml)	Bilirubin direct (mg/ml)	Bilirubin indirect (mg/ml)
Control	0.923 ± 0.022	0.146 ± 0.006	0.777 ± 0.026
Plf ext.	0.867 ± 0.060	0.145 ± 0.007	0.722 ± 0.061
TAA	2.280 ± 0.169	0.250 ± 0.024	2.031 ± 0.166
a	P = 0.0001	P = 0.0001	P = 0.0001
Plf ext+TAA	1.015 ± 0.033	0.184 ± 0.006	0.832 ± 0.028
a	P = 0.049	P = 0.0001	NS
b	P = 0.0001	P = 0.0001	P = 0.0001
γ radiation	1.307 ± 0.071	0.199 ± 0.014	1.103 ± 0.074
a	P = 0.0001	P = 0.0001	P = 0.0001
Plf ext+γ radiation	0.923 ± 0.033	0.162 ± 0.009	0.761 ± 0.030
a	NS	P = 0.020	NS
c	P = 0.0001	P = 0.0001	P = 0.0001
TAA+γ radiation	3.137 ± 0.076	0.281 ± 0.008	2.856 ± 0.079
a	P = 0.0001	P = 0.0001	P = 0.0001
b	P = 0.0001	P = 0.0001	P = 0.0001
c	P = 0.0001	P = 0.0001	P = 0.0001
Plf ext+TAA+γ radiation	1.310 ± 0.063	0.226 ± 0.005	1.084 ± 0.062
a	P = 0.0001	P = 0.0001	P = 0.0001
b	P = 0.0001	P = 0.0001	P = 0.0001
c	NS	P = 0.0001	NS
d	P = 0.0001	P = 0.0001	P = 0.009

Each value represents mean ± SD of 6 determinations.

a: significance over control group

b: significance over TAA group

c: significance over γ radiation group

d: significance over TAA+γ radiation group

NS: no significance

lipid hydroperoxides; CD: conjugated dienes; GSH: reduced glutathione; SOD: superoxide dismutase; Cat: catalase; AST: aspartateaminotransaminase; ALT: alanine aminotransferase; ALP: alkaline phosphatase; GGT: gamma-glutamyltransferase; ROS: reactive oxygen species; SD: standard deviation; NF-κB: nuclear factor-κB; TGF-β: tumour growth factor-beta; Abs₂₃₄: Absorbance at 234 nm.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

SZM designed the study, supervised the experiments, prepared Plf ext and wrote the manuscript. HEK performed the animal experiments. Both authors supervised the research assistants to carry out clinical chemistry assays. Both authors read and approved the final manuscript.

Acknowledgements

This study was financially supported by the National Centre for Radiation Research and Technology, Atomic Energy Authority, Cairo, Egypt.

References

- Pinzani M, Rombouts K: **Liver fibrosis: from the bench to clinical.** *Digestive Liver Dis* 2004, **36**:231-242.
- Hospers GA, Eisenhawer EA, de Vries EG: **The sulfhydryl containing compounds WR - 2721 and glutathione as radio - and chemoprotective agents. A review, indications for use and prospects.** *Br J Cancer* 1999, **80**:629-638.
- Porter WR, Neal RA: **Metabolism of thioacetamide and thioacetamide S-oxide by rat liver microsomes.** *Drug Metab Dispos* 1978, **6**:379-388.
- Hunter AL, Holscher MA, Neal RA: **Thioacetamide-induced hepatic necrosis: I. Involvement of the mixed-function oxidase enzyme system.** *J Pharmacol Exp Ther* 1977, **200**:439-448.
- Childs JFL, Siegler EA: **Uses of thioacetamide in agriculture.** *Science* 1945, **102**:68-72.
- Sanz N, Díez-Fernández C, Valverde AM, Lorenzo M, Benito M, Cascales M: **Malic enzyme and glucose-6-phosphate dehydrogenase gene expression increases in rat liver cirrhogenesis.** *Br J Cancer* 1997, **75**:487-492.
- Bruck R, Aeed H, Shirin H, Matas Z, Zaidel L, Avni Y, Halpern Z: **The hydroxyl radical scavengers dimethylsulfoxide and dimethylthiourea protect rats against thioacetamide-induced fulminant hepatic failure.** *J Hepatol* 1999, **31**:27-38.
- Barker EA, Smuckler EA: **Altered microsome function during acute thioacetamide poisoning.** *Mol Pharmacol* 1972, **8**:318-326.
- Muller D, Zimmerman SI, Schiller F: **Drug metabolism in rat liver injured by thioacetamide.** *Arch Toxicol* 1982, **5**:368-371.
- Johnson TS, El-Koraie AF, Skill NJ, Baddour NM, El Nahas AM, Nijloma M, Adam AG, Griffin M: **Tissue transglutaminase and the progression of human renal scarring.** *J Am Soc Nephrol* 2003, **14**(8):2052-2062.
- Mirza A, Liu SL, Frizell E, Zhu J, Maddukuri S, Martinez J, Davies P, Schwarting R, Norton P, Zern MA: **A role for tissue transglutaminase in hepatic injury and fibrogenesis, and its regulation by NF-kappa B.** *Am J Physiol* 1997, **272**(2):G281-288.
- Greenberg CS, Birckbichler PJ, Rice RH: **Transglutaminases: multifunctional cross-linking enzymes that stabilize tissues.** *FASEB J* 1991, **5**:3071-3077.
- Johnson TS, Griffin M, Thomas GL, Skill J, Cox A, Yang B, Nicholas B, Birckbichler PJ, Muchaneta-Kubara C, Meguid El Nahas A: **The role of transglutaminase in the rat subtotal nephrectomy model of renal fibrosis.** *J Clin Invest* 1997, **99**:2950-2960.

14. Aeschlimann D, Paulsson M: **Cross-linking of laminin-nidogen complexes by tissue transglutaminase: A novel mechanism for basement membrane stabilization.** *J Biol Chem* 1991, **266**:15308-15317.
15. Aeschlimann D, Paulsson M: **Transglutaminases: protein cross-linking enzymes in tissues and body fluids.** *Thromb Haemostasis* 1994, **71**:402-415.
16. Saralidze MA, Papava MB, Datunashvili IT, Sanikidze TV, Bakhtashvili VI: **Effectiveness of pliferon LB in gamma-radiotherapy.** *Georgian Med News* 2005, **124-125**:75-9.
17. Zhou D, Lauderback CM, Yu T, Brown SA, Butterfield DA, Thompson JS: **D609 inhibits ionizing radiation-induced oxidative damage by acting as a potent antioxidant.** *J Pharmacol Exp Ther* 2001, **298**:103-109.
18. Jagetia GC, Reddy TK: **Modulation of radiation-induced alteration in the antioxidant status of mice by naringin.** *Life Sci* 2005, **77(7)**:780-94.
19. Fadhel ZA, Amran S: **Effects of black tea extract on carbon tetrachloride-induced lipid peroxidation in liver, kidneys, and testes of rats.** *Phytother Res* 2002, **16(Suppl 1)**:S28-32.
20. Young SC, Wang CJ, Lin JJ, Peng PL, Hsu JL, Chou FP: **Protection effect of piper betel leaf extract against carbon tetrachloride-induced liver fibrosis in rats.** *Arch Toxicol* 2006, **81(1)**:45-55.
21. Christina AJM, Saraawathy GR, Heison Robert SJ, Kothai R, Chidambaranathan N, Nalini G, Therasal RL: **Inhibition of CCl4 induced liver fibrosis by Piper Longum Linn.** *Phytomedicine* 2006, **13**:196-198.
22. National Research Council: *Guide for the Care and Use of Laboratory Animals* 1985.
23. El Borai MS, Hessien M, El-key MM: **Effect of alpha-tocopherol on tissue transglutaminase and reversibility of thioacetamide-induced liver fibrosis in rats.** *Turk J Biochem* 2005, **31(1)**:13-20.
24. De Macedo P, Marrano C, Keillor JW: **A direct continuous spectrophotometric assay for transglutaminase activity.** *Anal Biochem* 2000, **285**:16-20.
25. Henry RJ: **Clinical Chemistry.** In *Principle and Techniques* Second edition. New York: Harper & Row; 1974:422-431.
26. Woessner JF Jr: **The determination of hydroxyproline in tissue and protein samples containing small proportions of this amino acid.** *Arch Biochem Biophys* 1961, **93**:440-447.
27. Beutler E, Duron O, Kelly BM: **Improved method of the determination of blood glutathione.** *J Lab Clin Med* 1963, **61(5)**:882-890.
28. Minami M, Yoshikawa H: **A simplified assay method of superoxide dismutase.** *Clinica Chimica Acta* 1979, **92**:337-342.
29. Sinha AK: **Colorimetric assay of catalase.** *Anal Biochem* 1972, **47**:389-394.
30. Yoshioka T, Kawada K, Shimada T, Mori M: **Lipid peroxidation in maternal and cord blood and protective mechanism against activated oxygen toxicity in the blood.** *Am J Obstet Gynecol* 1979, **135**:372-376.
31. Jiang ZK, Hunt JV, Wolf SP: **Detection of lipid hydroperoxides using Fox method.** *Anal Biochem* 1992, **202**:384-389.
32. Rechnagel RO, Gglende EA: **spectrophotometric detection of lipid conjugated dienes.** *Methods Enzymol* 1984, **105**:331-337.
33. Nowak D, Pierscinski G, Drzewoski J: **Ambroxol inhibits doxorubicin-induced lipid peroxidation in heart of mice.** *Free Radic Biol Med* 1995, **19**:659-663.
34. Teitz NW: **Fundamental of clinical chemistry.** *Chem Acta* 1976, **70**:602-609.
35. Reitman S, Frankel S: **A colorimetric method for the determination of serum glutamic oxalacetic and glutamic pyruvic transaminases.** *Amer J Clin Path* 1957, **28**:56-63.
36. Szasz G: **Reaction-rate method for gamma glutamyltransferase activity in serum.** *J Clin Chem* 1976, **22**:2051-2055.
37. Perry BV, Dumas BT, Bayse DD, Butler T, Cohen A, Fellows W, Garber CC, Howell B, Koch T, Krishnamurthy S, Louderback A, McComb RB, Miller D, Miller RR, Rand RN, Schaffer R: **A candidate reference method for determination of bilirubin in serum. Test for transferability.** *Clin Chem* 1983, **29(2)**:297-301.
38. Chen CS, Wu CH, Lai YC, Lee WS, Chen HM, Chen RJ, Chen LC, Ho YS, Wang YJ: **NF-kappaB-activated tissue transglutaminase is involved in ethanol-induced hepatic injury and the possible role of propolis in preventing fibrogenesis.** *Toxicology* 2008, **246(2-3)**:148-157.
39. Chen AP, Zhang L, Xu JY, Tang J: **The antioxidant (-)-Epigallocatechin-3-Gallate inhibits activated hepatic stellate cell growth and suppresses acetaldehyde induced gene expression.** *Biochem J* 2002, **368(3)**:695-704.
40. Iredale JP, Benyon RC, Pickering J, McCullen M, Northrop M, Pawley S, Hovell C: **Mechanisms of spontaneous resolution of rat liver fibrosis: hepatic stellate cell apoptosis and reduced hepatic expression of metalloproteinase inhibitors.** *J Clin Invest* 1998, **102**:538-549.
41. Lesort M, Tucholski J, Zhang J, Johnson GV: **Impaired mitochondrial function results in increased tissue transglutaminase activity in situ.** *J Neurochem* 2000, **75(5)**:1951-1961.
42. Sarkar S, Yadav P, Bhatnagar D: **Lipid peroxidative damage on cadmium exposure and alterations in antioxidant system in rat erythrocytes: A study with relation to time.** *Biometals* 1998, **11(2)**:153-157.
43. Devi PU, Ganasoundari A: **Modulation of glutathione and antioxidant enzymes by Ocimum sanctum and its role in protection against radiation injury.** *Indian J Exp Biol* 1999, **37**:262-268.
44. Erden Inal M, Kahraman A: **The protective effect of flavonol quercetin against ultraviolet A induced oxidative stress in rats.** *Toxicology* 2000, **154(1-3)**:21-29.
45. Abou-Seif MA, El-Naggar MM, El-Far M, Ramadan M, Salah N: **Prevention of biochemical changes in gamma-irradiated rats by some metal complexes.** *Clin Chem Lab Med* 2003, **41(7)**:926-933.
46. Oliinyk BV, Baraboi VA, Oliinyk SA, Horchakova NO: **Effect of splenoid on lipid peroxidation process and glutathione antioxidant system in rats exposed to fractionated radiation.** *Ukr Biokhim Zh* 2001, **73(1)**:73-77.
47. Ueda T, Toyoshima Y, Kushihashi T, Hishida T, Yasuhara H: **Effect of dimethyl sulfoxide pretreatment on activities of lipid peroxide formation, superoxide dismutase and glutathione peroxidase in the mouse liver after whole-body irradiation.** *J Toxicol Science* 1996, **18**:239-244.
48. Amonkar AJ, Nagabhushan M, D' Souza AV, Bhide SV: **Hydroxychavicol: a new phenolic antimutagen from betel leaf.** *Food Chem Toxicol* 1986, **24**:1321-1324.
49. Padma PR, Lalitha VS, Amonkar AJ, Bhide SV: **Anticarcinogenic effect of betel leaf extract against tobacco carcinogens.** *Cancer Lett* 1989, **45**:195-202.
50. Jeng JH, Kuo ML, Hahn LJ, Kuo MY: **Genotoxic and non-genotoxic effect of betel quid ingredients on oral mucosal fibroblasts in vitro.** *J Dent Res* 1994, **73**:1043-1049.
51. Choudhary D, Kale RK: **Antioxidant and non-toxic properties of piper betel leaf extract in vitro and in vivo studies.** *Phytother Res* 2002, **16**:461-466.
52. Lei D, Chan CP, Wang YJ, Wang TM, Lin BR, Huang CH, Lee JJ, Chen HM, Jeng JH, Chang MC: **Antioxidative and antiplatelet effect of aqueous inflorescence piper betel extract.** *J Agric Food Chem* 2003, **51**:2083-2088.
53. Jeng JH, Wang YJ, Chang WH, Wu HL, Li CH, Uang BJ, Kang JJ, Lee JJ, Hahn LJ, Lin BR, Chang MC: **Reactive oxygen species are crucial for hydroxychavicol toxicity toward KB epithelial cells.** *Cell Mol Life Sci* 2004, **61**:83-96.
54. Jalalpure SS, Patil MB, Prakash NS, Hemalata K, Manvi FV: **Hepatoprotective activity of the fruits of piper longum linn.** *Indian J Pharm Sci* 2003, **65(4)**:363-366.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

